

## Supporting Information

### Transcriptomics, proteomics and bioelectrochemical characterization of an exoelectrogen *Geobacter soli* grown with different electron acceptors

Xixi Cai<sup>1</sup>, Lingyan Huang<sup>1</sup>, Guiqin Yang<sup>1,\*</sup>, Zhen Yu<sup>2</sup>, Junlin Wen<sup>2</sup>, Shungui Zhou<sup>1</sup>

<sup>1</sup>Fujian Provincial Key Laboratory of Soil Environmental Health and Regulation, College of Resources and Environment, Fujian Agriculture and Forestry University, Fuzhou, China;

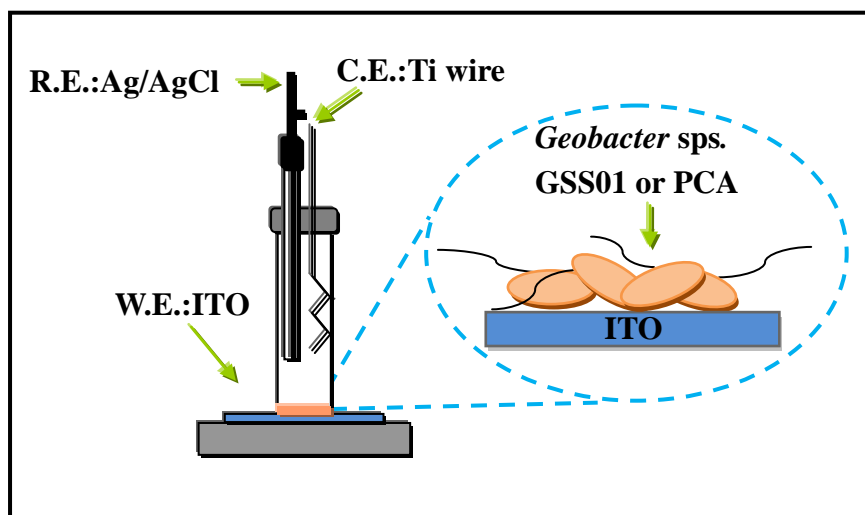
<sup>2</sup>Guangdong Key Laboratory of Integrated Agro-environmental Pollution Control and Management, Guangdong Institute of Eco-environmental Science & Technology, Guangzhou, China

**Running Head:** Extracellular electron transfer in *Geobacter*

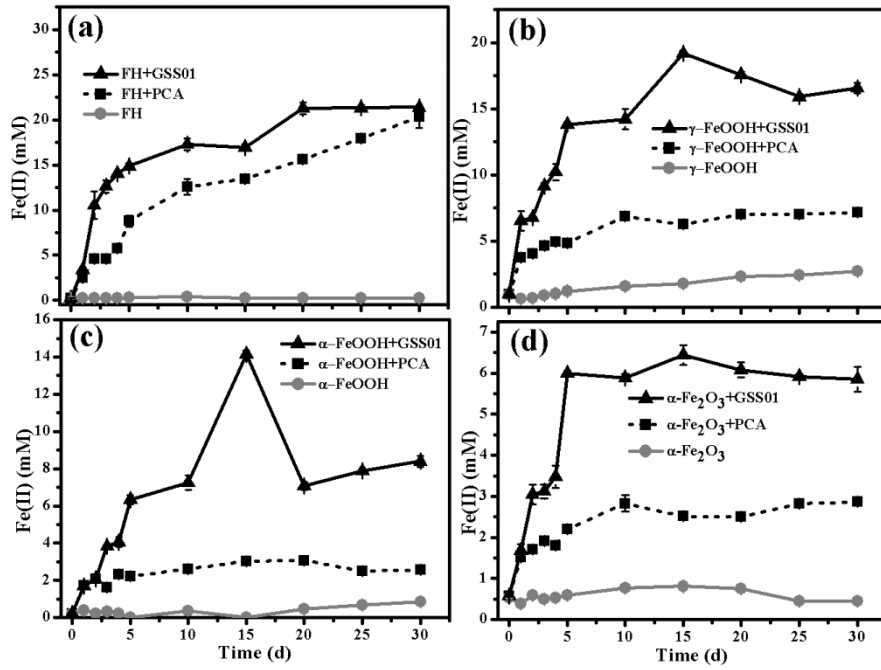
**\*Corresponding author:**

Guiqin Yang

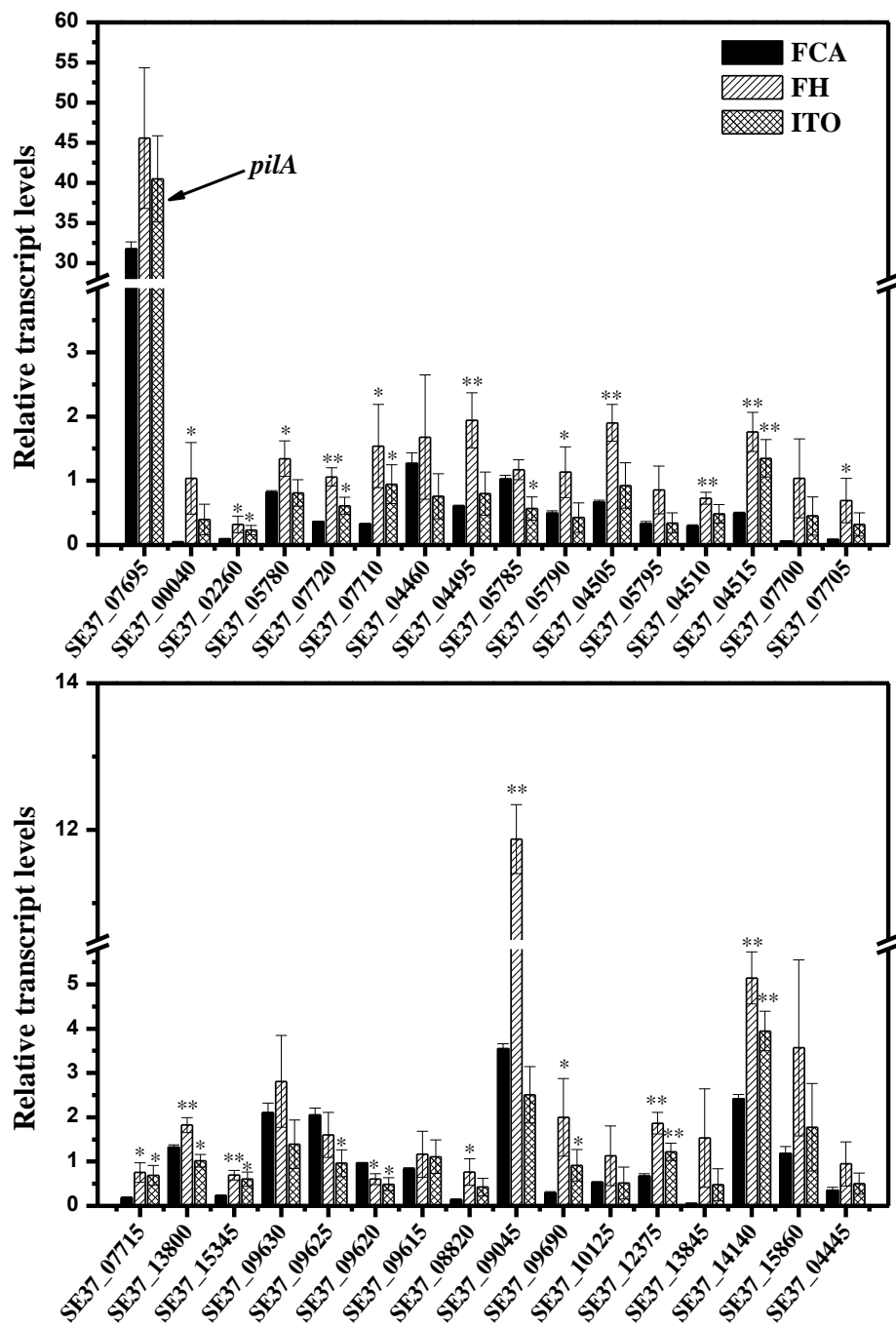
E-mail: ygqhappy@163.com



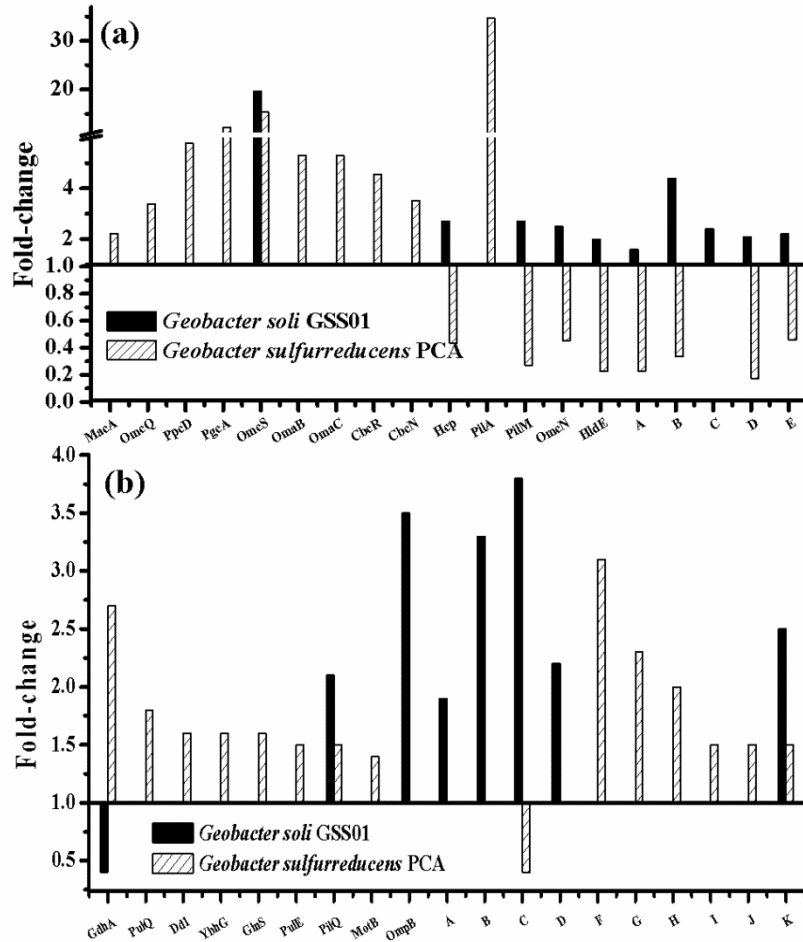
**Fig. S1** Diagram of a single-chamber, three-electrode system using ITO as working electrode (W.E: ITO), Ag/AgCl electrode as the reference electrode (R.E: Ag/AgCl) and Ti wire as the counter electrode.



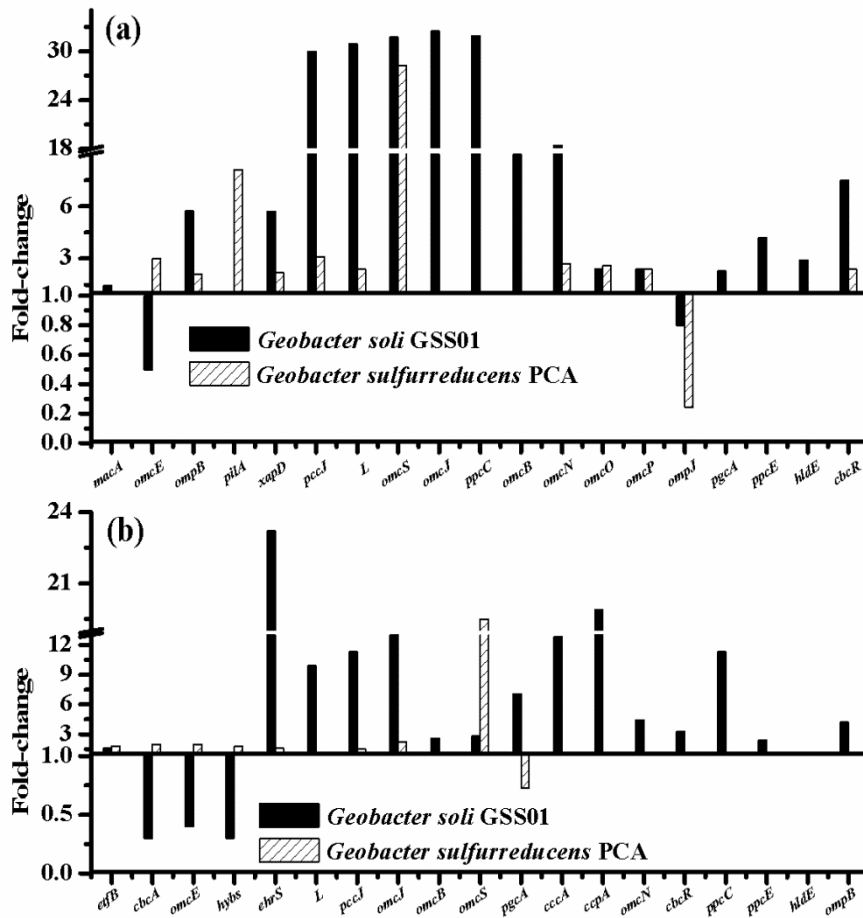
**Fig. S2** Total Fe(II) production from the reduction of 50 mM ferrihydrite (HFO), lepidocrocite ( $\gamma$ -FeOOH), goethite ( $\alpha$ -FeOOH) and hematite ( $\alpha$ -Fe<sub>2</sub>O<sub>3</sub>) by strain GSS01 and PCA.



**Fig. S3** Relative transcript levels of pilus-associated genes in *G. soli* cells grown with FH or electrode vs. FC. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$



**Fig. S4** Fold-change comparison of important proteins for electron transfer to Fe(III) oxides (a) or electrode (b) in *G. soli* and *G. sulfurreducens*. The data for *G. sulfurreducens* were obtained from Ding et al. (2008) and Kavanagh et al (2016). The y value was set as 1 for proteins that were not detected or were not significantly changed ( $P \geq 0.05$ ) in the proteomics analysis. Proteins that have no name were labeled with capital letters. The annotation of proteins in this figure is available in Table S5.



**Fig. S5** Comparison of transcript changes of important genes for electron transfer to Fe(III) oxides (a) or electrode (b) in *G. soli* and *G. sulfurreducens*. The data for *G. sulfurreducens* were obtained from Aklujkar et al. (2013) and Holmes et al (2006). The y value was set as 1 for genes that were not detected or were not significantly changed ( $P \geq 0.05$ ) in the transcriptomics analysis. Genes that have no name were labeled with capital letters. The annotation of genes in this figure is available in Table S5.

**Table S2** Selected genes with significantly differential abundance ( $P \leq 0.05$ ) in cells grown with FH or ITO in the transcriptomics analysis. -, unchanged or not find the gene names.

Locus ID	Gene name	Annotation	Difference (change fold)		
			FH vs. FC	ITO vs. FC	ITO vs. FH
<b>Proteins involved in metabolism and growth</b>					
<b>Tricarboxylic acid (TCA) cycle</b>					
SE37_02035	<i>aceF</i>	branched-chain alpha-keto acid dehydrogenase E2 subunit	11.2	-	-
SE37_07830	<i>korB</i>	2-oxoglutarate ferredoxin oxidoreductase subunit beta	1.7	0.6	0.3
SE37_03100	<i>pdhB</i>	pyruvate dehydrogenase E1 component subunit beta	1.6	-	-
SE37_10580	<i>acnA</i>	aconitate hydratase	2.0	-	0.6
SE37_14820	<i>pckA</i>	phosphoenolpyruvate carboxykinase	2.7	0.6	0.2
SE37_07825	<i>korC</i>	2-oxoglutarate:ferredoxin oxidoreductase subunit gamma	2.0	0.6	0.3
SE37_07845	<i>mdh</i>	malate dehydrogenase	0.6	0.5	-
SE37_09430	<i>gltA</i>	type I citrate synthase	0.2	0.2	-
SE37_07850	<i>icd</i>	isocitrate dehydrogenase	0.4	0.2	-
SE37_03070	<i>acn</i>	aconitate hydratase	0.7	0.6	-
SE37_09095	<i>frdB</i>	succinate dehydrogenase/fumarate reductase iron-sulfur subunit	0.2	0.1	0.6
SE37_09105	<i>frdC</i>	succinate dehydrogenase/fumarate reductase, cytochrome b558 subunit	0.1	0.1	-
SE37_09100	<i>frdA</i>	succinate dehydrogenase flavoprotein subunit	0.1	0.1	-
SE37_07835	<i>korA</i>	2-oxoglutarate ferredoxin oxidoreductase subunit alpha	-	0.5	0.3
SE37_07840	<i>korD</i>	2-oxoglutarate:ferredoxin oxidoreductase, ferredoxin subunit	-	0.5	0.5
SE37_06870	<i>acnB</i>	aconitate hydratase B	-	0.5	-
<b>Oxidative phosphorylation</b>					
SE37_14360	<i>buoF-2</i>	NADH dehydrogenase I subunit F	-	-	0.5
SE37_14370	<i>nuoE-1</i>	NADH dehydrogenase I subunit E	1.6	-	0.5
SE37_14375	<i>nuoBCD</i>	trifunctional NADH dehydrogenase I subunit B/C/D	-	0.7	-
SE37_14305	<i>nuoM-2</i>	NADH dehydrogenase I subunit M	7.8	5.5	-
SE37_13975	<i>atpA</i>	F0F1 ATP synthase subunit alpha	1.7	-	0.5
SE37_13985	<i>atpX</i>	ATP synthase F0, B' subunit	1.9	0.6	0.3
SE37_15965	<i>atpE</i>	ATP synthase F0 subunit C	1.7	-	-
SE37_13980	<i>atpH</i>	F0F1 ATP synthase subunit delta	1.8	0.7	0.4
SE37_16000	<i>nuoC</i>	NADH dehydrogenase I subunit C	-	0.5	-
SE37_16005	<i>nuoE</i>	NADH dehydrogenase I subunit D	-	0.5	-
SE37_16035	<i>nuoJ-1</i>	NADH dehydrogenase I subunit J	2.5	6.0	2.4
SE37_16055	<i>nuoN-1</i>	NADH dehydrogenase I subunit N	4.8	7.2	-
SE37_13990	<i>atpX</i>	ATP synthase F0, B' subunit	2.5	-	0.3
SE37_15305	<i>coxD</i>	cytochrome <i>c</i> oxidase, coo3-type, subunit IV	14.7	-	-
SE37_15300	<i>coxC</i>	cytochrome <i>c</i> oxidase, coo3-type, subunit III	9.4	-	-
SE37_15295	<i>coxA</i>	cytochrome <i>c</i> oxidase, coo3-type, cytochrome o subunit I	3.7	3.0	-
SE37_06970	<i>cydA</i>	cytochrome <i>bd</i> menaquinol oxidase, subunit I	0.7	0.6	-
SE37_01760	<i>hoxE</i>	bidirectional hydrogenase complex protein HoxE	-	0.4	0.3
SE37_01770	<i>hoxU</i>	bidirectional hydrogenase complex protein HoxU	-	0.5	0.5
SE37_01765	<i>hoxF</i>	bidirectional NAD-reducing hydrogenase, diaphorase subunit	-	0.6	0.4
SE37_15970	<i>atpB</i>	ATP synthase F0 subunit A	-	-	0.5
SE37_14380	<i>nuoA-2</i>	NADH dehydrogenase I subunit A	0.4	0.3	-
SE37_15990	<i>nuoA-1</i>	NADH dehydrogenase I subunit A	-	0.4	-
SE37_15995	<i>nuoB</i>	NADH dehydrogenase I subunit B	0.6	0.3	0.4
SE37_16050	<i>nuoM-1</i>	NADH dehydrogenase I subunit M	-	7.6	-
SE37_16040	<i>nuoK-1</i>	NADH dehydrogenase I subunit K	-	5.3	-
SE37_16045	<i>nuoL-1</i>	NADH dehydrogenase I subunit L	-	8.0	-

<b>Ribosomal proteins</b>					
SE37_11530	<i>rpsC</i>	30S ribosomal protein S3	6.5	4.6	-
SE37_11500	<i>rplE</i>	50S ribosomal protein L5	8.5	6.1	-
SE37_11685	<i>rplM</i>	50S ribosomal protein L13	1.6	-	0.5
SE37_05020	<i>rpsB</i>	30S ribosomal protein S2	2.0	-	0.3
SE37_01155	<i>rpsR</i>	30S ribosomal protein S18	2.1	0.6	0.3
SE37_01150	<i>rpsF</i>	30S ribosomal protein S6	1.7	0.5	0.3
SE37_03850	<i>rpmB</i>	50S ribosomal protein L28	1.7	-	0.6
SE37_01135	<i>rplY</i>	50S ribosomal protein L25/general stress protein Ctc	1.7	-	0.6
SE37_11540	<i>rpsS</i>	30S ribosomal protein S19	3.2	2.9	-
SE37_11535	<i>rplV</i>	50S ribosomal protein L22	6.1	4.5	-
SE37_11525	<i>rolP</i>	50S ribosomal protein L16	4.3	4.1	-
SE37_11515	<i>rpsQ</i>	30S ribosomal protein S17	7.2	6.1	-
SE37_11510	<i>rplN</i>	50S ribosomal protein L14	5.8	5.4	-
SE37_11600	<i>rplL</i>	50S ribosomal protein L7/L12	3.2	2.9	-
SE37_11505	<i>rplX</i>	50S ribosomal protein L24	4.3	4.7	-
SE37_11495	<i>rpsH</i>	30S ribosomal protein S8	4.0	4.5	-
SE37_11490	<i>rplF</i>	50S ribosomal protein L6	5.4	4.7	-
SE37_11485	<i>rplR</i>	50S ribosomal protein L18	3.5	4.4	-
SE37_11480	<i>rpsE</i>	30S ribosomal protein S5	2.9	3.7	-
SE37_11475	<i>rpmD</i>	50S ribosomal protein L30	12.4	8.9	-
SE37_11470	<i>rplO</i>	50S ribosomal protein L15	3.1	4.5	-
SE37_11430	<i>rplQ</i>	50S ribosomal protein L17	4.2	4.0	-
SE37_12715	<i>rpmE</i>	50S ribosomal protein L31	0.5	0.5	-
SE37_11580	<i>rpsG</i>	30S ribosomal protein S7	-	0.6	0.5
SE37_13245	<i>rplU</i>	50S ribosomal protein L21	-	0.5	0.4
SE37_13240	<i>rpmA</i>	50S ribosomal protein L27	-	0.5	0.3
SE37_02290	<i>rpsA</i>	30S ribosomal protein S1	-	0.6	0.6
SE37_03985	<i>rpsT</i>	30S ribosomal protein S20	-	0.6	0.5
SE37_12645	<i>rpsU-1</i>	30S ribosomal protein S21	-	0.6	0.4

### Proteins involved in EET

<b>Cytochromes</b>					
SE37_00350	<i>norC</i>	cytochrome <i>c</i>	19.1	8.5	-
SE37_00895	-	cytochrome <i>c</i>	3.3	2.1	-
SE37_01700	<i>omcQ</i>	cytochrome <i>c</i>	2.0	1.8	-
SE37_02825	<i>omcT</i>	cytochrome <i>c</i>	39.3	-	0.1
SE37_02835	-	cytochrome <i>c</i>	10.0	2.6	0.3
SE37_02870	<i>pccJ</i>	cytochrome <i>c</i>	30.0	11.3	0.4
SE37_03555	-	cytochrome <i>c</i>	15.6	25.3	-
SE37_03965	-	cytochrome <i>c</i>	3.3	1.5	0.5
SE37_05900	<i>pgcA</i>	cytochrome <i>c</i>	2.3	7.1	3.1
SE37_05905	<i>ppcE</i>	cytochrome <i>c</i>	4.2	2.4	-
SE37_06935	-	cytochrome <i>c</i>	4.3	3.3	-
SE37_08180	-	cytochrome <i>c</i>	3.1	3.9	-
SE37_08880	<i>omcI</i>	cytochrome <i>c</i>	4.6	2.1	0.5
SE37_11360	<i>cccA</i>	cytochrome <i>c</i>	9.2	12.8	-
SE37_11370	<i>ccpA</i>	cytochrome <i>c</i>	9.3	19.9	2.1
SE37_11705	-	cytochrome <i>c</i>	1.9	-	0.4
SE37_11760	<i>omcN</i>	cytochrome <i>c</i>	18.4	4.4	0.3
SE37_11765	-	cytochrome <i>c</i>	3.1	-	0.4
SE37_11830	<i>omcO</i>	cytochrome <i>c</i>	2.4	1.6	0.7
SE37_11835	<i>omcP</i>	cytochrome <i>c</i>	2.4	-	0.4
SE37_11905	-	cytochrome <i>c</i>	3.1	-	-
SE37_11920	<i>cbcR</i>	cytochrome <i>c</i>	7.5	3.3	-
SE37_11940	-	cytochrome <i>c</i>	1.9	2.8	-
SE37_11945	-	cytochrome <i>c</i>	2.0	-	0.4

SE37_13340	<i>imcH</i>	cytochrome <i>c</i>	2.1	-	-
SE37_14185	-	cytochrome <i>c</i>	3.9	2.9	-
SE37_14295	-	cytochrome <i>c</i>	11.9	-	-
SE37_15065	-	cytochrome <i>c</i>	3.1	-	0.4
SE37_15310	-	cytochrome <i>c</i>	7.9	-	-
SE37_15520	<i>cbcL</i>	cytochrome <i>c</i>	7.6	-	0.5
SE37_16115	<i>ppcB</i>	cytochrome <i>c</i>	1.7	0.3	0.2
SE37_16120	<i>ppcC</i>	cytochrome <i>c</i>	31.9	11.3	-
SE37_01425	<i>omcJ</i>	cytochrome <i>c</i>	32.5	17.9	-
SE37_02820	<i>omcS</i>	cytochrome <i>c</i>	31.8	2.8	0.1
SE37_02865	-	cytochrome <i>c</i>	30.9	-	0.3
SE37_01725	<i>omcB</i>	cytochrome <i>c</i>	9.0	2.6	0.3
SE37_00770	-	cytochrome <i>c</i>	0.7	-	-
SE37_00775	<i>obcC</i>	cytochrome <i>c</i>	0.4	0.4	-
SE37_00785	-	cytochrome <i>c</i>	0.3	0.3	-
SE37_00905	<i>omcE</i>	cytochrome <i>c</i>	0.5	0.4	-
SE37_01435	-	cytochrome <i>c</i>	0.3	0.7	2.7
SE37_02090	-	cytochrome <i>c</i>	0.4	0.1	0.4
SE37_02785	-	cytochrome <i>c</i>	0.2	0.1	0.6
SE37_05760	-	cytochrome <i>c</i>	0.6	0.6	-
SE37_08440	-	cytochrome <i>c</i>	-	-	0.5
SE37_00575	-	cytochrome <i>c</i>	-	-	0.6
SE37_00875	-	cytochrome <i>c</i>	-	-	0.7
SE37_09635	-	cytochrome <i>c</i>	-	-	0.4
SE37_11335	-	cytochrome <i>c</i>	0.4	0.6	1.6
SE37_11345	-	cytochrome <i>c</i>	0.5	0.3	0.5
SE37_11955	-	cytochrome <i>c</i>	0.4	1.5	4.3
SE37_00200	<i>macA</i>	cytochrome <i>c</i>	1.4	1.6	-
SE37_01170	<i>omcX</i>	cytochrome <i>c</i>	-	1.9	-
SE37_07370	-	cytochrome <i>c</i>	-	3.9	-
SE37_09795	<i>ppcD</i>	cytochrome <i>c</i>	-	2.0	1.8
SE37_12935	<i>nrfA</i>	cytochrome <i>c</i>	-	4.6	-
SE37_12940	<i>nrfH</i>	cytochrome <i>c</i>	-	2.8	-
SE37_16085	-	cytochrome <i>c</i>	-	1.5	1.9
SE37_04285	<i>omcZ</i>	cytochrome <i>c</i>	-	0.5	-
SE37_08660	-	cytochrome <i>c</i>	-	0.4	0.4
SE37_11710	-	cytochrome <i>c</i>	-	0.5	0.5
SE37_12855	-	cytochrome <i>c</i>	-	0.7	0.5
<b>Pili</b>					
SE37_00040	<i>pilB</i>	pilus assembly protein PilB	24.6	-	-
SE37_02260	<i>pilB</i>	pilus assembly protein PilB	3.5	2.6	-
SE37_05780	<i>pilB</i>	pilus assembly protein PilB	1.6	-	-
SE37_05785	<i>pilM</i>	pilus assembly protein PilM	-	0.6	0.5
SE37_07720	<i>pilB</i>	pilus assembly protein PilB	2.9	1.7	-
SE37_07710	<i>pilC</i>	pilus assembly protein PilC	4.7	2.9	-
SE37_04495	<i>pilM</i>	pilus assembly protein PilM	3.2	-	0.4
SE37_05790	<i>pilN</i>	pilus assembly protein PilN	2.3	-	-
SE37_04505	<i>pilO</i>	pilus assembly protein PilQ	2.9	-	0.5
SE37_04510	<i>pilP</i>	pilus assembly protein PilP	2.5	-	-
SE37_04515	<i>pilQ</i>	pilus assembly protein PilQ	3.5	2.7	-
SE37_07705	<i>pilS</i>	pilus assembly protein PilS	8.3	-	-
SE37_07715	<i>pilT</i>	pili twitching motility protein PilT	4.1	3.8	-
SE37_15345	<i>pilT</i>	pili twitching motility protein PilT	3.1	2.7	-
SE37_08820	<i>pilZ</i>	pilus assembly protein PilZ	5.5	-	-
SE37_09045	<i>pilZ</i>	pilus assembly protein PilZ	3.3	-	-
SE37_09690	<i>pilZ</i>	pilus assembly protein PilZ	6.8	3.1	-
SE37_12375	<i>pilZ</i>	pilus assembly protein PilZ	2.8	1.8	0.7

SE37_13800	<i>pilT</i>	pili twitching motility protein PilT	-	-	0.6
SE37_14140	<i>pilZ</i>	pilus assembly protein PilZ	2.1	1.6	0.8
SE37_09620	<i>pilX</i>	pilus assembly protein PilX	0.6	0.5	-
SE37_09625	<i>pilW</i>	pilus assembly protein PilW	-	0.5	-
<b>Other EET genes</b>					
SE37_04245	<i>hldE</i>	bifunctional heptose 7-phosphate kinase/heptose 1-phosphate adenylyltransferase	2.9	-	0.4
SE37_08195	<i>ompB</i>	laccase	5.7	4.2	-
SE37_12120	-	ligand-gated channel	35.7	13.8	-
SE37_10395	-	ligand-gated channel	4.9	2.3	-
SE37_11970	-	hypothetical protein	0.2	-	3.0
SE37_11965	-	porin	0.2	-	4.7

**Table S3** Selected proteins with significantly differential abundance ( $P \leq 0.05$ ) in cells grown with FH or ITO in the proteomics analysis. -, unchanged or not find the gene names.

Locus ID	Gene name	Annotation	Difference (change fold)	
			FH vs. FC	ITO vs. FC
<b>Proteins involved in metabolism and cell growth</b>				
<b>TCA cycle</b>				
SE37_14820	<i>pckA</i>	phosphoenolpyruvate carboxykinase	0.6	-
SE37_09430	<i>gltA</i>	type I citrate synthase	0.4	0.4
SE37_10580	<i>acnA</i>	aconitate hydratase	2.5	2.4
SE37_11700	<i>icd</i>	3-isopropylmalate dehydrogenase	2.7	-
SE37_09945	<i>fumB</i>	fumarate hydratase	0.2	0.6
SE37_05525	<i>asl</i>	L-aspartate oxidase	0.6	-
SE37_06870	<i>acnB</i>	aconitate hydratase B	-	0.4
<b>Oxidative phosphorylation</b>				
SE37_15995	<i>nuoB</i>	NADH-quinone oxidoreductase subunit B	2.3	-
SE37_16005	<i>nuoE</i>	NADH dehydrogenase	1.8	-
SE37_16015	<i>nuoF-1</i>	NADH dehydrogenase	0.1	0.7
SE37_13965	<i>atpB</i>	ATP synthase subunit beta	3.3	-
SE37_13980	<i>atpH</i>	ATP synthase FOF1 subunit delta	2.3	-
SE37_13970	<i>atpG</i>	ATP FOF1 synthase subunit gamma	0.3	-
SE37_16020	<i>nuoG</i>	NADH dehydrogenase	-	0.5
SE37_00450	-	NADH-ubiquinone oxidoreductase subunit 3	-	0.6
<b>Ribosomal proteins</b>				
SE37_11440	<i>rpsD</i>	30S ribosomal protein S4	0.04	0.3
SE37_09005	<i>rpsA</i>	30S ribosomal protein S1	0.6	0.4
SE37_11495	<i>rpsH</i>	30S ribosomal protein S8	0.5	0.2
SE37_01150	<i>rpsF</i>	30S ribosomal protein S6	0.4	0.4
SE37_05020	<i>rpsB</i>	30S ribosomal protein S2	0.3	0.2
SE37_11680	<i>rpsI</i>	30S ribosomal protein S9	0.1	0.2
SE37_11530	<i>rpsC</i>	30S ribosomal protein S3	0.1	0.4
SE37_11445	<i>rpsK</i>	30S ribosomal protein S11	0.1	0.3
SE37_11450	<i>rpsM</i>	30S ribosomal protein S13	0.1	0.1
SE37_11540	<i>rpsS</i>	30S ribosomal protein S19	0.1	0.2
SE37_11685	<i>rplM</i>	50S ribosomal protein L13	0.1	0.2
SE37_11505	<i>rplX</i>	50S ribosomal protein L24	0.1	0.3
SE37_11635	<i>rpmG</i>	50S ribosomal protein L33	0.1	-
SE37_11605	<i>rplJ</i>	50S ribosomal protein L10	0.3	0.3
SE37_07210	<i>rplG</i>	50S ribosomal protein L7	0.3	-
SE37_11510	<i>rplN</i>	50S ribosomal protein L14	0.2	0.4
SE37_11610	<i>rplA</i>	50S ribosomal protein L1	0.2	0.2
SE37_11550	<i>rplW</i>	50S ribosomal protein L23	0.1	0.2
SE37_11470	<i>rplO</i>	50S ribosomal protein L15	0.1	0.6
SE37_11560	<i>rplC</i>	50S ribosomal protein L3	0.1	0.2
SE37_07150	<i>rpmF</i>	50S ribosomal protein L32	0.1	-
SE37_01165	<i>rpli</i>	50S ribosomal protein L9	0.1	0.2
SE37_11500	<i>rplE</i>	50S ribosomal protein L5	0.07	0.3
SE37_01040	<i>rplS</i>	50S ribosomal protein L19	0.05	0.3
SE37_11490	<i>rplF</i>	50S ribosomal protein L6	0.05	0.2
SE37_01015	<i>rpsP</i>	30S ribosomal protein S16	-	0.2
SE37_02290	<i>rpsA</i>	30S ribosomal protein S1	-	0.5
SE37_12645	<i>rpsU-1</i>	30S ribosomal protein S21	-	0.5
SE37_11520	<i>rpmC</i>	50S ribosomal protein L29	-	0.3
SE37_11615	<i>rplK</i>	50S ribosomal protein L11	-	0.3
SE37_13245	<i>rplU</i>	50S ribosomal protein L21	-	0.2

SE37_11555	<i>rplD</i>	50S ribosomal protein L4	-	0.2
SE37_01135	<i>rplY</i>	50S ribosomal protein L25	-	0.5
SE37_11600	<i>rplL</i>	50S ribosomal protein L7/L12	-	0.5
SE37_11685	<i>rplM</i>	50S ribosomal protein L13	-	0.2
SE37_11505	<i>rplX</i>	50S ribosomal protein L24	-	0.3
SE37_11605	<i>rplJ</i>	50S ribosomal protein L10	-	0.3
SE37_11510	<i>rplN</i>	50S ribosomal protein L14	-	0.4
SE37_11610	<i>rplA</i>	50S ribosomal protein L1	-	0.2
SE37_11550	<i>rplW</i>	50S ribosomal protein L23	-	0.2
SE37_11470	<i>rplO</i>	50S ribosomal protein L15	-	0.6
SE37_11560	<i>rplC</i>	50S ribosomal protein L3	-	0.2
SE37_01165	<i>rplI</i>	50S ribosomal protein L9	-	0.2
SE37_01040	<i>rplI</i>	50S ribosomal protein L19	-	0.3
SE37_11490	<i>rplF</i>	50S ribosomal protein L6	-	0.2

### Proteins involved in EET

<b>Cytchromes</b>				
SE37_02820	<i>omcS</i>	cytochrome C	19.7	-
SE37_11760	<i>omcN</i>	cytochrome C	2.5	-
SE37_04285	<i>omcZ</i>	cytochrome C	0.3	0.2
SE37_15520	<i>cbcL</i>	cytochrome C	0.2	-
SE37_00785	<i>nrfB</i>	cytochrome C	0.2	-
SE37_11705	-	cytochrome C	0.2	0.2
SE37_13340	<i>imcH</i>	cytochrome C	0.2	-
SE37_12935	<i>nrfA</i>	cytochrome C	0.1	-
SE37_04675	-	cytochrome C	0.1	-
SE37_08660	-	cytochrome C	0.1	-
SE37_01170	<i>omcX</i>	cytochrome C	0.1	-
SE37_01700	-	cytochrome C	0.1	-
SE37_05900	<i>pgcA</i>	cytochrome C	-	0.2
SE37_11710	-	cytochrome C	-	0.3
<b>Pili proteins</b>				
SE37_04495	<i>pilM</i>	pilus assembly protein PilM	2.7	-
SE37_04515	<i>pilQ</i>	pilus assembly protein PilQ	2.1	2.1
SE37_07715	<i>pilT</i>	type IV pili twitching motility protein PilT	0.6	-
SE37_02260	<i>pilB</i>	pilus assembly protein PilB	0.5	-
SE37_07710	<i>pilC</i>	pilus assembly protein PilC	0.4	-
SE37_04505	<i>pilO</i>	pilus assembly protein PilO	0.2	-
<b>Other EET proteins</b>				
SE37_04245	<i>hldE</i>	bifunctional heptose 7-phosphate kinase/heptose 1-phosphate adenylyltransferase	2.0	-
SE37_11965	-	porin	4.4	3.3
SE37_08195	<i>ompB</i>	laccase		
SE37_12120	-	ligand-gated channel	1.6	1.9
SE37_10790	-	membrane protein	2.4	3.8
SE37_11970	-	hypothetical protein	2.1	2.2
SE37_10395	-	ligand-gated channel	2.2	2.0

**Table S5** Annotation of genes in Fig. S4 and Fig. S5

<b>Gene name</b>	<b>annotation</b>	<b>Locus ID in strain PCA</b>	<b>Locus ID in strain GSS01</b>	<b>Amino acid similarity</b>
omcS	Cytochrome <i>c</i>	GSU2504	SE37_02820	94.7%
omcB	Cytochrome <i>c</i>	GSU2737	SE37_01745	83.8%
omcE	Cytochrome <i>c</i>	GSU0618	SE37_00905	93.5%
xapD	ABC transporter, ATP-binding protein	GSU1501	SE37_07675	59.5%
macA	Cytochrome <i>c</i> peroxidase	GSU0466	SE37_00200	95.7%
ompB	Laccase family multicopper oxidase	GSU1394	SE37_08195	94.9%
pccJ	Cytochrome <i>c</i>	GSU2494	SE37_02870	97.9%
omcJ	Cytochrome <i>c</i>	GSU0701	SE37_01425	98.1%
ppcC	Cytochrome <i>c</i>	GSU0365	SE37_16120	93.7%
ppcD	Cytochrome <i>c</i>	GSU1024	SE37_09795	97.8%
ppcE	Cytochrome <i>c</i>	GSU1760	SE37_05905	91.1%
pgcA	Lipoprotein cytochrome <i>c</i>	GSU1761	SE37_05900	88.1%
omcN	Lipoprotein cytochrome <i>c</i>	GSU2898	SE37_11760	98.8%
omcO	Cytochrome <i>c</i>	GSU2912	SE37_11830	98.5%
omcP	Cytochrome <i>c</i>	GSU2913	SE37_11835	97.1%
omcQ	Cytochrome <i>c</i>	GSU0592	SE37_00775	97.6%
ompJ	Outer membrane channel	GSU3304	SE37_13550	54.3%
ccdA	Cytochrome <i>c</i> biogenesis protein	GSU2085	SE37_04245	96.1%
cbcR	Methaquinol oxidoreductase complex, lipoprotein cytochrome <i>c</i> subunit	GSU2930	SE37_11920	94.0%
hybS	Ni/Fe hydrogenase, small subunit	GSU0782	SE37_10925	98.7%
ehrS	NAD-dependent dehydrogenase subunit	GSU0745	SE37_01620	96.8%
cccA	Cytochrome B6	GSU2811	SE37_11360	95.9%
ccpA	Cytochrome <i>c</i> peroxidase	GSU2813	SE37_11370	98.0%
omaB	Cytochrome <i>c</i>	GSU2732	SE37_01740	97.4%
omaC	Cytochrome <i>c</i>	GSU2738	SE37_01720	97.4%
cbcN	Methaquinol oxidoreductase complex Cbc6, cytochrome <i>c</i> subunit	GSU2934	SE37_11940	98.1%
hcp	Iron-sulfur-oxygen hybrid cluster protein	GSU0674	SE37_01195	98.5%
PilA	Geopilin domain 1 protein	GSU1496	SE37_07695	85.3%
pilM	Type IV pilus biogenesis ATPase	GSU2032	SE37_04495	97.4%
pilQ	Type IV pilus secretin lipoprotein	GSU2028	SE37_04515	87.2%
gdhA	Glutamate dehydrogenase	GSU1305	SE37_08570	98.4%
pulQ	Type II secretion system secretin lipoprotein	GSU1778	SE37_05805	92.5%
pulE	Type II secretion system ATPase	GSU1783	SE37_05780	96.9%
ddl	D-alanine-D-alanine ligase	GSU3066	SE37_12520	95.6%
ybhG	Efflux pump, RND family, membrane fusion protein	GSU2823	SE37_11390	95.7%
glnS	Glutamine-tRNA ligase	GSU3366	SE37_14915	98.4%
motB	Peptidoglycan-binding lipoprotein, OmpA family	GSU1013	SE37_09840	96.9%
A	Ligand-gated channel, TonB-dependent copper receptor	GSU2982	SE37_12120	96.7%
B	Outer-membrane porin protein	GSU2939	SE37_11965	94.8%
C	Outer-membrane channels, porin superfamily	GSU0810	SE37_10790	92.1%
D	Hypothetical protein	GSU2940	SE37_11970	69.6%
E	Ligand-gated channel, porin superfamily	GSU0883	SE37_10395	82.5%
F	Glutamate synthase, FMN-Fe(II)-binding domain protein	GSU1239	SE37_08825	99.8%
G	FAD-dependent pyridine nucleotide-disulfide oxidoreductase	GSU1237	SE37_08835	97.5%
H	Efflux pump, RND family, membrane protein	GSU0496	SE37_00385	98.8%
I	Transcriptional regulator, MarR family	GSU2362	SE37_03395	93.0%
J	Phosphoglucosyltransferase/phosphomannosyltransferase family protein	GSU2013	SE37_04590	96.6%
K	Amino acid aminotransferase	GSU0117	SE37_13945	96.1%
L	Cytochrome <i>c</i>	GSU2495	SE37_02865	97.4%

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