

Supplementary Material

Supplementary Table 1: Antibodies and fluorochrome combinations used in the flow cytometry panels

Antibody	Fluorochrome /Channel	Clone	Supplier	Volume in 50 μ L
CD4 panel				
Live/Dead Dye	BV510	Aqua fluorescent reactive dye	Invitrogen	0.5
CD4	BV785	SK3	BioLegend	0.6
HLA-DR	PE	L243	BioLegend	1
CD3	FITC	DREG-56	BioLegend	1
CD8	APC-Cy7	SK1	BioLegend	2
Granzyme B	BV421	GB11	BioLegend	4
TNF- α	PE-Cy7	MAb11	BioLegend	1
IFN- γ	Alexa Flour700	B27	BioLegend	0.5
Monocyte panel				
NIR	APC-Cy7	Near Infrared Dye	Invitrogen	1
CD14	PE-Cy7	M5E2	ELabscience	2,5
CD3	ECD	A07748	Beckman Coulter	1,2
HLA-DR	BV605	L243	BioLegend	0.5
TNF- α	APC	R101	Sinobiological	2.5
IL-6	FITC	MQ2-13A5	ELabscience	1.2
IL-1 β	PE	AS10	BD	0.6

Supplementary information

BCG culture

Bacillus Calmette-Guerin (BCG) is the only licensed vaccine for TB and is a TB strain made up of live *Mycobacterium bovis*. BCG shares several proteins with Mtb but lacks genes from the region of interest (RD-1) which confer virulence to Mtb (23).

We cultured BCG stocks to exponential growth phase in 7H9 media supplemented with glycerol, tween-80, and Oleic Albumin Dextrose Catalase (OADC). Once cultures reached the exponential growth phase (OD value between 0.4 and 0.7), stocks were made by storing the cells in 7H9 media with 15% glycerol and transferring cells to -80°C for long-term storage. Frozen stocks were used to determine colony forming units (CFU) by plating different dilutions of thawed BCG in 7H10 agar plates and incubating plates at 37°C. Cells were counted after 3 weeks of incubation. BCG was used at MOI of 1.

H37Rv culture

Mtb H37Rv was cultured in the BSL-3 laboratory in 7H9 media supplemented with glycerol, tween 80, and OADC until culture reached exponential growth phase. Clumps were dispersed using glass beads and bacterial stocks were prepared by storing cells in

7H9 media supplemented with 15% glycerol. To determine CFUs, different dilutions of thawed bacterial stocks were cultured in 7H10 agar plates and incubated at 37°C for 3 weeks. Colonies were manually counted to determine CFUs. H37Rv was used at MOI of 1.

List of cytokines used in Luminex and bead regions

APRIL [52], BAFF [65], BLC (CXCL13) [15], CD30 [34], ENA-78 (CXCL5) [55], Eotaxin-2 (CCL24) [37], Eotaxin-3 (CCL26) [77], FGF-2 [75], Fractalkine (CX3CL1) [39], IL-16 [47], IL-2R (CD25) [22], IL-20 [26], I-TAC (CXCL11) [46], MCP-2 (CCL8) [18], MCP-3 (CCL7) [66], MDC (CCL22) [30], MIF [67], MIG (CXCL9) [54], TNF-RII [45], TRAIL (CD253) [29], TSLP [61], TWEAK [35], CD40L (CD154) [74], Eotaxin (CCL11) [33], Gro-alpha (CXCL1) [61], G-CSF (CSF-3) [42], GM-CSF [44], HGF [46], IFN alpha [48], IP-10 (CXCL10) [22], IFN gamma [43], IL-1 alpha [62], IL-1 beta [18], IL-2 [19], IL-3 [73], IL-4 [20], IL-5 [21], IL-6 [25], IL-7 [26], IL-8 (CXCL8) [27], IL-9 [52], IL-10 [28], IL-12p70 [34], IL-13 [35], IL-15 [65], IL-17A (CTLA-8) [36], IL-18 [66], IL-21 [72], IL-22 [76], IL-23 [63], IL-27 [14], IL-31 [37], LIF [15], M-CSF [67], MCP-1 (CCL2) [51], MIP-1 alpha (CCL3) [12], MIP-1 beta (CCL4) [47], MIP-3 alpha (CCL20) [56], MMP-1 [64], NGF beta [55], SDF-1 alpha (CXCL12) [13], SCF [39], TNF alpha [45], TNF beta [54], VEGF-A [78]