

Table S1 Bacterial strains and plasmids used in this study

Strain or plasmid	Genotype or properties	Reference
<i>S. maltophilia</i>		
KJ	A clinical <i>S. maltophilia</i> isolate	Hu et al. 2008
KJΔFur	<i>S. maltophilia</i> KJ mutant of <i>fur</i> gene; Δ <i>fur</i>	Shih et al. 2022
KJΔHemP	<i>S. maltophilia</i> KJ mutant of <i>hemP</i> gene; Δ <i>hemP</i>	Shih et al. 2022
KJΔEnt	<i>S. maltophilia</i> KJ mutant of <i>entF</i> and <i>entA</i> genes; Δ <i>entF</i> , Δ <i>entA</i>	Liao et al. 2020
KJΔEntΔHemA	<i>S. maltophilia</i> KJΔEnt mutant of <i>hemA</i> gene; Δ <i>entF</i> , Δ <i>entA</i> , Δ <i>hemA</i>	Shih et al. 2022
KJΔEntΔ3896	<i>S. maltophilia</i> KJΔEnt mutant of <i>smlt3896</i> gene; Δ <i>entF</i> , Δ <i>entA</i> , Δ <i>smlt3896</i>	This study
KJΔEntΔHemU	<i>S. maltophilia</i> KJΔEnt mutant of <i>hemU</i> gene; Δ <i>entF</i> , Δ <i>entA</i> , Δ <i>hemU</i>	This study
KJΔEntΔTonB2	<i>S. maltophilia</i> KJΔEnt mutant of <i>tonB2</i> gene; Δ <i>entF</i> , Δ <i>entA</i> , Δ <i>tonB2</i>	This study
KJΔEntΔTonB1	<i>S. maltophilia</i> KJΔEnt mutant of <i>tonB1</i> gene; Δ <i>entF</i> , Δ <i>entA</i> , Δ <i>tonB1</i>	This study
KJΔEntΔTonB1ΔTonB2	<i>S. maltophilia</i> KJΔEnt mutant of <i>tonB1</i> and <i>tonB2</i> gene; Δ <i>entF</i> , Δ <i>entA</i> , Δ <i>tonB1</i> , Δ <i>tonB2</i>	This study
<i>E. coli</i>		
DH5α	F- φ80d/ <i>acZAM15</i> Δ(<i>lacZYA-argF</i>) <i>U169 deoR recA1 endA1 hsdR17</i> (r _K ⁻ m _K ⁺) <i>phoA supE44λ thi-1 gyrA96 relA1</i>	Invitrogen
S17-1	λ <i>pir</i> ⁺ mating strain	Simon et al. 1986

Plasmids		
pEX18Tc	<i>sacB oriT</i> , Tc ^r	Hoang et al. 1998
pRK415	Mobilizable broad-host-range plasmid cloning vector, RK2 origin; Tc ^r	Keen et al. 1988
pΔ3896	pEX18Tc with an internal-deleted <i>smlt3896</i> gene; Tc ^r	This study
pΔHemU	pEX18Tc with an internal-deleted <i>hemU</i> gene; Tc ^r	This study
pΔTonB1	pEX18Tc with an internal-deleted <i>tonB1</i> gene; Tc ^r	This study
pΔTonB2	pEX18Tc with an internal-deleted <i>tonB2</i> gene; Tc ^r	This study
pHemU	pRK415 with an intact <i>hemU</i> gene; Tc ^r	This study
pTonB1	pRK415 with an intact <i>tonB1</i> gene; Tc ^r	This study

Reference

- Hoang, T. T., Karkhoff-Schweizer, R. R., Kutchma, A. J., and Schweizer, H. P. (1998). A broad-host-range Flp-FRT recombination system for site-specific excision of chromosomally-located DNA sequences: application for isolation of unmarked *Pseudomonas aeruginosa* mutants. *Gene*. 212, 77-86.
- Hu, R. M., Huang, K. J., Wu, L. T., Hsiao, Y. J., and Yang, T. C. (2008). Induction of L1 and L2 β-lactamases of *Stenotrophomonas maltophilia*. *Antimicrob Agents Chemother*. 52, 1198-1200.
- Keen, N. T., Tamaki, S., Kobayashi, D., and Trollinger, D. (1988). Improved broad-host-range plasmids for DNA cloning in gram-negative bacteria. *Gene*. 70, 191-197.
- Liao, C. H., Chen, W. C., Li, L. H., Lin, Y. T., Pan, S. Y., and Yang, T. C. (2020). AmpR of *Stenotrophomonas maltophilia* is involved in stenobactin synthesis and enhanced β-lactam resistance in an iron-depleted condition. *J. Antimicrob. Chemother*. 75, 3544–3551.
- Shih, Y. L., Wu, C. M., Lu, H. F., Li, L. H., Lin, Y. T., and Yang, T. C. (2022). Involvement of the *hemP-hemA-smlt0796-smlt0797* operon in hemin acquisition by *Stenotrophomonas maltophilia*. *Microbiol Spectr*. 10, e0032122.
- Simon, R., O'Connell, M., Labes, M., and Puhler, A. (1986). Plasmid vector for the genetic analysis and manipulation of *Rhizobia* and other Gram-negative bacteria. *Methods Enzymol*. 118, 640-659.